# GUIDELINES ON STRANGLES

## **GUIDELINES ON STRANGLES**

#### The Disease

Strangles is a disease of the lymph nodes of the equine upper respiratory tract. It is caused by the bacterium *Streptococcus equi* (S. equi) and is endemic within the horse population of the United Kingdom.

#### **Notification Procedures**

There are no legal notification requirements for strangles in the UK, although it is advisable to inform the national breeders' associations if infection occurs. Under the Rules of Racing (Section C30 Duty to report communicable diseases), racehorse trainers are obliged to report likely or confirmed strangles to the British Horseracing Authority (BHA) when it occurs among horses in training.

### **Clinical Signs**

Affected horses typically have a high temperature, cough, poor appetite, nasal discharge and swollen or abscessed lymph nodes of the head, which can appear as open sores. Some infected horses may become very ill and the disease may become fatal if the bacterium spreads to other parts of the body ('bastard strangles') or the respiratory tract is occluded by swollen lymph nodes (hence the term 'strangles'). However, in some horses, a nasal discharge without glandular swelling is sometimes all that is seen.

### Transmission of Disease

Direct contact between infected horses is the most obvious means of transmitting the infection but the hands and equipment of staff, farriers or veterinary surgeons can spread it indirectly. The bacterium is discharged (shed) from draining abscesses and the nose, and it may survive in the environment, particularly in water troughs. Good hygiene is therefore essential in controlling the disease. The incubation period (the time between infection occurring and clinical signs developing) is usually about one week but may be longer. Horses incubating the disease may shed *S. equi* before the onset of obvious clinical signs and so may spread the infection to in-contacts before the first case becomes apparent.

A small but important proportion of horses that have recovered from strangles become persistently infected (most commonly in their guttural pouches) with *S. equi* for months or even years. These 'carriers' are less susceptible to reinfection, and they





may have no obvious clinical signs of disease but can intermittently shed *S. equi*, which can then infect naive horses. These subclinical carriers are probably the most important factor in persistence of infection on premises between outbreaks and can initiate new outbreaks following their inadvertent movement to new premises.

#### **Prevention**

More details on methods for prevention of introduction of strangles onto equine premises are available in the 'Strategy to eradicate and prevent Strangles (STEPS)' document (https://ww1.sruc.ac.uk/media/5v3jl2yg/steps-guidelines.pdf)

Ideally, all horses entering any stud or stable premises should be quarantined for a period of 3-4 weeks and monitored closely for any clinical signs of an infectious disease, including daily temperature monitoring, particularly in the period immediately after arrival. Any horse that develops a nasal discharge or other signs consistent with strangles should be isolated and tested for the presence of, or exposure to, S. equi.

The strangles blood test can be used to identify horses that have elevated antibody responses to *S. equi* and are therefore likely to have been exposed to this pathogen in the recent past, enabling the identification of potentially infectious animals before or immediately after movement. A further blood test at the end of the quarantine process can be used to identify animals that may have seroconverted since their arrival, consistent with recent exposure to *S. equi*. It is recommended that any quarantine batches of horses that include seropositive animals, as well as those seroconverting whilst in quarantine, not be released until their infectious status has been shown to be negative for presence of *S. equi* (see Diagnosis below).

Veterinary advice should be sought to determine whether use of vaccination may be appropriate on the basis of a specific risk assessment and bearing in mind that the modified-live vaccine may trigger positive results in diagnostic tests for strangles. See Appendix 8 for information on available vaccines.

### **Diagnosis**

Strangles is diagnosed either directly by detection of *S. equi* itself or indirectly by detection of rising levels of antibody against *S. equi* in blood samples, although presence of antibodies against *S. equi* does not necessarily indicate that an animal is still infectious to other horses. Direct detection of *S. equi* is either by laboratory isolation or by qPCR detection of its DNA from nasopharyngeal swabs, abscess contents and/ or guttural pouch washes or contents (empyema/chondroids). It should be noted that low bacterial numbers, the concurrent presence of the closely related *S. zooepidemicus* or recent antibiotic treatment, may make the detection of *S. equi* by culture more difficult and less sensitive than qPCR.

When taking nasopharyngeal swabs, it is particularly important to sample the back of the pharynx around the opening of the guttural pouch, using



specially designed elongated swabs with enlarged absorbent heads (see 1 below). There is no need to use smaller, guarded swabs as the main purpose of swabbing for strangles is to optimise the chances of detecting the organism if it is present. Shedding of *S. equi* into the nasopharynx often occurs intermittently, so repeated swabbing is recommended to confirm negative results. *S. equi* should be more reliably confirmed or excluded following testing by qPCR alone or by culture and qPCR of frank pus from obvious draining abscesses, nasal discharges or guttural pouch washes or contents (empyema or chondroids).

The carrier state (continued presence of *S. equi* infection in the absence of clinical signs) may be diagnosed or excluded by sequential nasopharyngeal swabs or, preferably, endoscopic examination ('scoping') of the guttural pouches and submission of guttural pouch washes or contents (empyema or chondroids) for testing by qPCR alone or by culture and qPCR. A series of three nasopharyngeal swabs, usually collected one week apart, will result in detection, by positive qPCR, on at least one of the swabs in >90% of carrier horses. As the sensitivity of *S. equi* detection for identifying guttural pouch carriers on three nasopharyngeal swabs is broadly equivalent to testing bilateral guttural pouch samples, the latter approach is the recommended sampling protocol for determining infectious status in seropositive, healthy horses.

Although carriers only shed *S. equi* intermittently, most carriers maintain specific antibodies in their blood and these antibodies can be detected by a blood ELISA test, which may provide a useful tool to help identify some, but not all, carrier animals. Recent investigations of the blood ELISA in detecting chronic carriers have highlighted that negative blood ELISA results, either as single or paired samples, do not guarantee absence of a carrier state. It is therefore preferable for all potential carriers irrespective of their serological status, especially those of high or unknown risk status, to be examined by guttural pouch endoscopy and sampling with screening for *S. equi* by qPCR alone or by culture and qPCR.

#### **Control of Infection**

More details on methods for control and eradication of strangles on equine premises are available in the 'Strategy to eradicate and prevent Strangles (STEPS)' document, which is accessible at

https://ww1.sruc.ac.uk/media/5v3jl2yq/steps-guidelines.pdf.

The spread of *S. equi* may be limited by the early detection of shedders among newly affected horses and their in-contacts by appropriate testing (see above). Any suspected cases should be isolated immediately.

Young and elderly horses are most susceptible to infection and should be monitored closely. All infected horses and their in-contacts should remain in strict isolation, under the direction of the attending veterinary surgeon, and with the highest possible standards of hygiene.

Regular disinfection of water troughs should be performed in order to minimise the infectious dose that in-contact horses receive and so reduce the severity of disease.

Horses should not enter affected premises unless they can be kept in strict isolation from all possible sources of infection. No infected or in-contact animal should be released from isolation or veterinary supervision until they have been tested conclusively negative for active shedding and the carrier state, as described above.

#### **Treatment**

The treatment of horses with clinical signs of strangles using antibiotics remains controversial and any essential treatment will be determined by the attending veterinary surgeon, who will be best placed to consider all relevant risks. For further advice regarding the treatment of carrier horses please consult the attending vet.

#### Confirmation of Freedom from Disease

Shedding of S. equi usually ends rapidly after complete recovery but may continue intermittently for several weeks after clinical signs have resolved in some carrier horses. Therefore, no convalescent horse or in-contact horses can be considered free from infection until either three negative nasopharyngeal swabs have been obtained or the horse has been tested negative on bilateral guttural pouch samples. Negative results for S. equi by qPCR alone or by culture and qPCR indicate freedom from infection and the carrier state in the large majority of cases, but not all, so vigilance must be maintained. In deciding on the best time to commence testing to confirm freedom from infection after an outbreak of strangles it should be noted that this is likely to be a trade-off between starting sooner and finding a proportion of convalescing horses that continue to harbour S. equi that would if left longer have cleared the infection naturally and starting later and identifying fewer true subclinical S. equi carriers that require treatment and re-testing. Experience suggests that the best compromise is for clearance testing to commence at least four weeks after the last clinical signs of strangles have been observed.

# Testing horses prior to their introduction to new premises

Horses entering new premises should be quarantined for 3-4 weeks in case they are incubating diseases such as equine influenza or strangles, infected horses may or may not have clinical signs or they may be subclinical carriers. All new arrivals should be examined for signs of illness (high temperatures, dullness, not eating, nasal discharge, swollen or abscessed lymph glands around the head or neck). Any horses with such signs should be immediately isolated and veterinary advice sought.

Routine use of the strangles ELISA blood test during isolation can identify previously infected and potentially infectious horses quickly. Ideally samples should be taken on arrival and after three weeks isolation to check for rising antibody levels (seroconversion) indicating an immune response after exposure to *S. equi*. If any of the quarantined horses are ELISA blood test positive on either the first or second test then further swab/wash testing (using qPCR or culture and

qPCR) is required in order to determine whether the positive animals are carrying *S. equi* (see Diagnosis above).

#### **Export Certification**

Strangles is not notifiable by law. However, no horse with clinical signs or recent contact with this disease should be exported.

# Testing horses at the end of a strangles outbreak

Following an outbreak of strangles, the best time to detect a carrier horse is a minimum of 30 days after the last clinical signs are seen. Shedding usually ends rapidly after recovery although it may continue intermittently in some horses.

# Further information for veterinary surgeons

Detection of *Steptococcus* equi subspecies equi using a triplex qPCR assay: https://www.sciencedirect.com/science/article/pii/S1090023312003103

Strategy to eradicate and prevent Strangles (STEPS): https://ww1.sruc.ac.uk/media/5v3jl2yq/steps-guidelines.pdf

Combining two serological assays optimizes sensitivity and specificity for the identification of Streptococcus equi subsp. equi exposure: https://www.sciencedirect.com/science/article/pii/S1090023313000567

Streptococcus equi infections: current best practice in the diagnosis and management of 'strangles':

https://www.magonlinelibrary.com/doi/full/10.12968/ukve.2021.5.2.S.3

Streptococcus equi Infections in Horses: Guidelines for Treatment, Control, and Prevention of Strangles—Revised Consensus Statement: https://onlinelibrary.wiley.com/doi/full/10.1111/jvim.15043